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Exploration and Collection of Cucumber Mosaic Virus Isolates of Horticultural Plants from Bali



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Abstract

Information regarding the existence and biological character of CMV isolates from several horticultural plants from Bali is still limited. The results of this research on information are very much needed in the further research and development of science and technology, due to it is known that CMV isolates from different hosts can cause different symptoms of infection or specific symptoms in certain plants. Based on these problems, this research was conducted through exploration, collection, and biological testing of CMV isolates found in several horticultural plants in Bali. Therefore, the existence and biological characteristics of each isolate were known. The results showed that all isolates obtained from the field, as many as 36 isolates, only 18 isolates showed symptoms of N. glutinousa. The isolates showed symptoms in N. glutinosa plants were isolates from cucumber plants 4 isolates, eggplant 3 isolates, pumpkin 1 isolate, 3 chili isolates, 2 isolates tomatoes, 2 beans isolates, and watermelons 3 isolates. The types of symptoms that appeared varied, i.e., 10 symptomatic mosaic isolates, 4 malformed isolates, and 4 necrotic symptomatic isolates. Detection of I-ELISA using CMV antiserum succeeded in getting 6 positive CMV isolates, i.e., isolates derived from cucumber plants (CMV-MB isolates), eggplant (CMV-TRB isolates) watermelon (CMV-SB isolates), chili (CMV-CB isolates), pumpkin (CMV-LB isolates), and tomatoes (CMV-TB isolates). The results of biological testing of CMV isolates from Bali, mosaic symptoms occurred in the indicator plants Nicotiana glutinosa, Capsicum annuum (big chili), Cucumis sativus (cucumber), Lycopersum esculentum (tomato), and Cucurbita moschata (pumpkin). Severe mosaic symptoms only occurred in Cucumis sativus (cucumber) plants. In Vigna sinensis (long bean) infection CMV-Bali isolates only cause symptoms of malformation and even cause no symptoms.

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1. Introduction

Cucumber Mosaic Virus (CMV) is a species of virus originating from genus Cucumovirus, family Bromoviridae (Mochizuki and Ohki, 2012). It is round shape with 30 nm diameter, has four types of nucleic acids, which is a single-threaded RNA (Palukaitis et al., 1992; Fauquet et al., 2005). CMV is also one of the mosaic disease-causing viruses found in Cucurbitaceae plants. CMV can infect 1200 species in more than 100 families of vegetable and horticulture plants. These characteristics make CMV a wrong plant virus is economically important in commercial plants (Palukaitis et al., 1992). Important economic plants can be infected by CMV are big chili, cayenne pepper, paprika, tomato, cucumber, melon, banana, eggplant, chrysanthemum, lilies, and a number of other ornamental plants (Agrios, 2005).

CMV has a wide host range, its spread in all countries and strains with different biological properties have been reported from various places. CMV has many strains and differences between strains depend on the host type, the symptoms produced, the transmission nature, and other characteristics (Francki *et al.*, 1979; Agrios 2005). The disease symptoms due to CMV infection vary depending on the host species or CMV strain. Therefore, it appears to be a character of the host reaction to CMV infection. Pandawani *et al.*, (2017), in cucumber plants (*Cucumis sativus*) and other *Cucurbitaceae* members, CMV infection causes mosaic and dwarf symptoms and reduces fruit quantity and quality. In tomato plants (*Lycopersicon esculentum*), CMV infections cause mosaic, dwarf, fern leaf reduction (McNab *et al.*, 1983) symptoms. In *Capsicum annuum*, CMV infections cause mosaic symptoms, blistering, fruit malformations, and together with other viruses (TMV, TEV, PVY, and CVMV) cause symptoms of complex viral infections (curly symptoms).

2. Materials and Methods

Exploration of host samples infected viruses

Virus exploration is conducted on several types of symptomatic horticultural plants infected with the virus. Samples were taken from 9 regencies in Bali, included Jembrana, Buleleng, Tabanan, Badung, Gianyar, Bangli, Klungkung, Karangasem, and Denpasar City. Host plants suspected of being infected with CMV were collected and taken at least three samples from one type of plant in each location. To obtain a variety of isolates with more samples collected were samples that showed different symptoms or virus-specific symptoms, namely mosaic, blistering, necrotic leaves/fruit, and leaf malformations.

Propagation of CMV isolates from different hosts

Propagation of isolates is conducted through mechanical inoculation of symptomatic host saplings infected with the virus. Mechanical viral inoculation was conducted 1 gram of CMV inoculum plant leaves were crushed in sterile mortal by adding 10 ml of phosphate/phosphate buffer (1:10 b/v) containing 1% β -mercaptoethanol (0, 01 M, pH 7.0). The *sap yan* is formed immediately inoculated on *N. glutinosa* plant which is on the surface of the first leaf which has been fully opened, the leaves of which have been sprinkled with carborundum 600 mesh first. The plants that have been inoculated are kept in insect vectors that are resistant to symptoms of the disease. The same symptomatic plants on the CMV isolate source plant in the field were then selected, for further purification of CMV isolates.

Purification of CMV isolates

Virus purification was conducted through biological testing to select CMV isolates. The plants needed to obtain pure CMV isolates were *Nicotiana glutinosa*, *Cucumis sativus*, and *Chenopodium amaranticolor*. The purification *Pandawani*, N. P., *Javandira*, C., & *Hanum*, F. (2018). Exploration and collection of cucumber mosaic virus isolates of horticultural plants from Bali. International Research Journal of Engineering, IT & Scientific Research, 4(6), 44-54. https://doi.org/10.21744/irjeis.v4n6.340

method of CMV isolates was carried out as follows: a sample from the field that had been propagated in N. glutinosa (the result of point 2.2) was infected with *C. sativus* in the cotyledon section, and then infected with *C. amaranticolor* (repeated three times). Furthermore, the virus obtained was re-inoculated into *N. glutinosa* and *C. sativus* plants. Furthermore, the obtained isolates were maintained on *N. glutinosa* and to ensure that CMV isolates were obtained, serological testing of isolates with the *Indirect Enzyme-linked immunosorbent assay* was performed (I-ELISA) using serum anti-CMV (AGDIA, USA).

CMV isolates collection through the biological test on indicator plants

Indicator plants used to study the symptoms of infection by all isolates obtained were plants: *C. annuum* (big chili), *Cucumis sativus* (cucumber), *L. esculentum* (tomato), *Solanum melongena* (green eggplant), *Vigna sinensis* (long beans), and *Cucurbita moschata* (pumpkin). All CMV positive isolates were inoculated on several indicator plants. The differences in isolates are expected to be identified based on differences in symptoms in indicator plants so that CMV isolates collection from several hosts was obtained.

3. Results and Discussions

3.1 Collection of plants infected with viruses in the field

Isolates collection, as well as symptoms observations that occur in plants, suspected being infected with CMV have been conducted and data obtained are shown in Table 1. Isolates were obtained from the field as many as 36 isolates namely derived from cucumber plants (5 isolates), big chili (6 isolates), tomatoes (4 isolates), long beans (4 isolates), watermelon (4 isolates), pumpkin (4 isolates) and melons (4 isolates).

Table 1
Isolates collection and host symptoms suspected being infected with CMV

Origin of isolate Regency	Host plants Symptoms host field		Sample Code	
1. Bangli				
	cucumber	mosaic	1 M	
	long beans	necrotic	2 K	
	eggplant	mosaic	3 Tr	
	pumpkin	Malformation	4 L	
2. Amlapura	•		·	
	watermelon	malformation	5 S	
	cucumber	Malformation	6 M	
	pumpkin	mosaic	7 L	
	melon	necrotic	8 Me	
3. Semarapura	•		·	
	tomato	malformation	9 T	
	chili	mosaic	10 C	
	eggplant	necrotic	11 Tr	
	watermelon	mosaic	12 S	
4. Gianyar	•		· 	
	cucumber	necrotic	13 M	
	pumpkin	mosaic	14 L	
	eggplant	Malformation	15 Tr	
	tomato	mosaic	16 T	
5. Singaraja				
	melon	mosaic	17 Me	
	chili	malformation	18 C	

mosaic

19 Tr

eggplant

	long beans	mosaic	20 K
6. Jembrana			
	cucumber	malformation	21 M
	watermelon	necrotic	22 S
	melon	mosaic	23 Me
	chili	necrotic	24 C
7. Tabanan			
	tomato	malformation	25 T
	cucumber	mosaic	26 M
	pumpkin	necrotic	27 L
9. D. 1	chili	mosaic	28 C
8. Badung	melon	magaia	29 Me
	chili	mosaic motel	30 C
	long beans	malformation	31 K
	tomato	malformation	31 K 32 T
9. Denpasar	tomato	manormation	32 1
y. Denpasai	chili	necrotic	33 C
	watermelon	mosaic	34 S
	eggplant	mosaic	35 Tr
	long beans	motel	36 K
A. Long beans	B. Cuo	cumber	C. Tomato
D. Eggplant	E. P	umpkin	F. Chili

Figure 1. Symptoms of plants in the field suspected being infected with CMV

The plants infected with the virus in the field show complex symptoms, which are infected with several viruses simultaneously. The sample obtained it is expected that there is a single infection by CMV. Symptoms appear in the host plants in the field are suspected of being infected with CMV are very varied or different, ranging from mild to

severe mosaic, blistering, and leaf malformations (Figure 1). The different symptoms in the same host plant are expected to be obtained by different CMV strains.

3.2 Purification results of CMV isolates

The plants are from the field suspected being infected with the virus then purified/conducted purification of the virus with biological techniques to separate CMV from other viruses, using specific indicator plants. All isolates obtained from the field were as many as 36 isolates, after purification only 18 isolates showed symptoms in *N. glutinousa* plants, 18 other isolates did not show symptoms. Isolates who showed symptoms were 10 mosaic symptom isolates, 4 isolates from leaf malformations and 4 isolates from necrotic symptoms. The isolates showing symptoms of *N. glutinosa* were isolates from cucumber plants 4 isolates, eggplant 3 isolates, pumpkin 1 isolate, chili 3 isolates, tomatoes 2 isolates, beans 2 isolates, and watermelons 3 isolates. The incubation period in *N. glutinosa* plants ranged from 11 days to 15 days after inoculation (Table 2).

Table 2
Type of symptoms in *N. glutinosa* indicator plants infected with CMV isolates from different hosts

No.	Sample	Original host	Incubation	Symptoms in indicator
	Code	field	period (hsi)	plants/N. glutinosa
1	1 M	cucumber	12	mosaic
2	2 K	long beans	14	necrotic
3	3 Tr	eggplant	13	mosaic
4	5 S	watermelon	11	malformation
5	6 M	cucumber	11	malformation
6	10 C	chili	14	mosaic
7	11 Tr	eggplant	14	necrotic
8	13 M	cucumber	12	necrotic
9	14 L	pumpkin	13	mosaic
10	16 T	tomato	14	mosaic
11	19 Tr	eggplant	13	mosaic
12	20 K	long beans	15	mosaic
13	22 S	watermelon	13	mosaic
14	24 C	chili	15	malformation
15	25 T	tomato	12	malformation
16	26 M	cucumber	11	mosaic
17	33 C	chili	15	necrotic
18	34 S	watermelon	12	mosaic

CMV infects systemically in many plants. Older organ or tissue plants that develop before being infected with the virus are usually not affected by the virus presence, but the young tissue or cells that develop after being infected with the virus are very affected and generally show acute symptoms. Virus symptoms will increase several days after the infection, then decrease to a certain extent or until the plant dies.

3.3 Serology test results of CMV isolates

The variation in symptoms caused by CMV will be very difficult to identify only based on the symptoms. In addition, it is also difficult to distinguish other CMV from *Cucumovirus* isolates (such as *Alfalfa mosaic virus*, *Tomato aspermy virus*, *and Peanut stunt virus*). To further ensure, the virus obtained is CMV, the isolates that have been successfully purified are then tested serologically using I-ELISA test. The isolated sample was expressed positively if the results of the absorbance measurements at 405 nm wavelength with ELISA reader (Table 3) had a value twice as large as the negative control value (Matthews, 2002).

Table 3
I-ELISA absorption value at A 405 nm of CMV isolates

No.	Sample Code	Origin of the host	I-ELISA / Antiserum		
	_		CMV / absorbance value		
1	1 M	cucumber	2.524		
2	2 K	long beans	0.522		
3	3 Tr	eggplant	2.356		
4	5 S	watermelon	2.584		
5	6 M	cucumber	0.620		
6	10 C	chili	2.422		
7	11 Tr	eggplant	0.426		
8	13 M	cucumber	0.524		
9	14 L	pumpkin	2.482		
10	16 T	tomato	0.465		
11	19 Tr	eggplant	0.540		
12	20 K	long beans	0.582		
13	22 S	watermelon	0.488		
14	24 C	chili	0.522		
15	25 T	tomato	2.528		
16	26 M	cucumber	0.642		
17	33 C	chili	0.584		
18	34 S	watermelon	0.634		
	Buffer		0.238		
	Positive control	·	2.634		
	Negative control		0.466		

Detection by I-ELISA using CMV antiserum succeeded in getting 6 positive CMV isolates (Figure 2) derived from cucumber plants (CMV-MB isolates), eggplant plants (CMV-TRB isolates) watermelon plants (CMV-SB isolates), chili plants (CMV-CB isolates, pumpkin plants (CMV-LB isolates) and tomato plants (CMV-TB isolates) (Table 4). Other isolates also showed symptoms of *N. glutinosa* appeared to be from CMV I-ELISA test result is not positive.

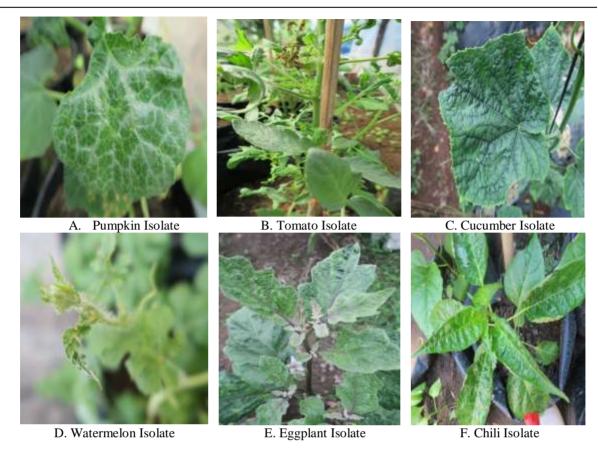


Figure 2. CMV-Bali isolates

Table 4 CMV positive isolates from different hosts

No.	Sample	Origin of host	Symptom type on	I-ELISA	Isolate name
	code		indicator plants/N.		of CMV Bali
			glutinosa		
1	1 M	Cucumber	Mosaic	+	CMV-MB
2	3 Tr	eggplant	Malformation	+	CMv-TRB
3	5 S	Watermelon	Malformation	+	CMV-SB
4	10 c	Chili	Mosaic	+	CMV-CB
5	14 L	Pumpkin	Mosaic	+	CMV-LB
6	25 T	Tomatoes	Malformation	+	CMV-TB

3.4 Indicators plant symptoms on the biological test of CMV isolates

The symptoms result from infection with six CMV isolates from Bali on several important indicator plants are highly varied and some are asymptomatic. It appears varying from 6 days after inoculation to 13 days after inoculation with severe mosaic, mosaic, local malformations, and necrotic symptoms (Table 5).

Table 5
Symptoms of indicator plants infected with CMV isolates from Bali

	CMV-Bali Isolates							
Indicator plant	CMV-MB	CMV-TRB	CMV-SB	CMV-CB	CMV-LB	CMV-TB		
	Symptoms							
Nicotiana glutinosa	mosaic	malformation	mosaic	mosaic	malformation	mosaic		
Capsicum annuum (big chili)	mosaic	mosaic	malformation	mosaic	mosaic	malformation		
Cucumis sativus	weight	mosaic	weight	asymptomatic	mosaic	mosaic		
(cucumber),	mosaic		mosaic					
Lycopersum.								
esculentum	malformation	malformation	mosaic	mosaic	malformation	malformation		
(tomato),								
Vigna sinensis	malformation	asymptomatic	asymptomatic	asymptomatic	asymptomatic	malformation		
(long beans),								
Cucurbita								
moschata	mosaic	asymptomatic	malformation	mosaic	mosaic	malformation		
(pumpkin)								
Chenopodium	necrotic	necrotic	necrotic	necrotic	necrotic	necrotic		
amaranticolor								

In indicator plants, *Nicotiana glutinosa isolates* CMV-MB, CMV-SB, CMV-CB, and CMV-TB cause mosaic symptoms, whereas other isolates cause symptoms of malformation. In *Capsicum annuum* (big chili) isolates CMV-MB, CMV-TRB, CMV-CB and CMV-LB cause symptomatic mosaics, whereas CMV-SB and CMV-TB isolate cause symptoms of malformation. In *Cucumis sativus* (cucumber) plants, CMV-MB isolates and CMV-SB showed severe mosaic symptoms, CMV-TRB, CMV-LB, and CMV-TB isolates caused mosaic symptoms while CMV-CB isolates showed no symptoms. In *Lycopersum esculentum* plants (tomato), CMV-SB and CMV-CB isolate because of mosaic symptoms while CMV-MB, CMV-TRB, CMV-LB, and CMV-TB isolates cause symptoms of malformation. In indicator plants of *Vigna sinensis* (long bean) isolates CMV-MB and CMV-TB cause symptoms of malformation while CMV-TRB isolates, CMV-SB, CMV-CB, and CMV-LB do not cause symptoms. In the *Cucurbita moschata* plant (pumpkin), CMV-MB, CMV-CB, and CMV-LB isolate caused mosaic symptoms, CMV-SB and CMV-TB isolates caused symptoms of malformation while CMV-TRB isolates caused symptoms. In *Chenopodium amaranticolor* plants all isolates cause local necrotic symptoms.

Severe mosaic symptoms only occur in *Cucumis sativus* (cucumber) plants due to infection with CMV-MB isolates and CMV-SB. Mosaic symptoms occur in indicator plants *Nicotiana glutinosa* (due to infection with CMV-MB, CMV-SB, CMV-CB, and CMV-TB isolates), in *Capsicum annuum* plants (big chili) (due to infection with CMV-MB isolates, CMV-TRB), CMV-CB and CMV-LB, in *Cucumis sativus* (cucumber) plants, (due to CMV-TRB, CMV-LB and CMV-TB isolates), in *Lycopersum esculentum* (tomato) plants (due to CMV-SB and CMV isolates) -CB), in *Cucurbita moschata* (pumpkin) plants (due to infection with CMV-MB, CMV-CB and CMV-LB isolates) In *Vigna sinensis* (long bean) infection CMV-Bali isolates did not cause mosaic symptoms, only caused symptoms of malformations (due to infection with CMV-MB and CMV-TB isolates) and even CMV isolates from Bali, namely CMV-TRB isolates, CMV-SB, CMV-CB, and CMV-LB did not cause symptoms.

4. Conclusion

1. Exploration of all isolates from the field as many as 36 isolates, and only 18 isolates showed symptoms in *N. glutinousa* plants. The isolates showed symptoms in *N. glutinosa* plants were isolates from cucumber plants 4 isolates, eggplant 3 isolates, pumpkin 1 isolate, 3 chili isolates, 2 isolate tomatoes, 2 bean isolates, and 3 watermelon isolates. The symptoms types appear vary, 10 of which are symptomatic mosaic, 4 are symptomatic malformations, and 4 necrotic symptomatic isolates.

2. Detection of I-ELISA using CMV antiserum succeeded in getting 6 CMV positive isolates namely isolates derived from cucumber plants (CMV-MB isolates), eggplant (CMV-TRB isolates) watermelon (CMV-SB isolate), chili (CMV-CB isolate), pumpkin (CMV-LB isolate) and tomatoes (CMV-TB isolates).

3. Biological test of CMV isolates from Bali, mosaic symptoms occur in the indicator plant *Nicotiana glutinosa* included *Capsicum annuum* (big chili), *Cucumis sativus* (cucumber), *Lycopersum esculentum* (tomato), and *Cucurbita moschata* (punpkin). Heavy mosaic symptoms only occur on *Cucumis sativus* (cucumber) plant. In *Vigna sinensis* (long bean) plant infection, CMV-Bali isolates only cause symptoms of malformation and do not even cause symptoms.

Conflict of interest statement and funding sources

The authors declared that they have no competing interest. The study was financed by Directorate of Research and Community Service, Ministry of Research, Technology, and Higher Education of the Republic of Indonesia.

Statement of authorship

The authors have a responsibility for the conception and design of the study. The authors have approved the final article.

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